Hong Kong Institute of Medical Laboratory Sciences

Quality Assurance Programme

August Survey (2026)

Anatomical Pathology

HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES

QUALITY ASSURANCE PROGRAMME

Anatomical Pathology

INSTRUCTIONS

1. All tests should be performed as soon as possible after the receipt of the specimens.
2. Unless otherwise specified, all survey material had been fixed in 10% buffered formalin and paraffin embedded.
3. The survey material should be treated, as far as possible, like a patient specimen.
4. The survey material should be treated as potentially hazardous and standard laboratory safety precautions should be taken as usual in your laboratory.
5. To complete the survey, you should send back the stained slides together with appropriate controls, and the questionnaire for statistical analysis and compilation of results.
6. Do NOT forward any particulars of your laboratory except your assigned confidential Laboratory Code. **Late return will be counted as not done and the results will NOT be accepted for assessment and statistical analysis.**
7. All stained slides should be properly labelled with the assigned 4-digit laboratory code but not the identity of your laboratory.
8. Place the labelled slides inside the slide mailers. Please ensure the mountant is completely dried before placing the slides inside the slide mailer.
9. Return the slide mailers and completed questionnaire in the enclosed foam-padded envelope.
10. Arrange return of survey material with the courier service provider according to the instruction as per attached before on **20 August 2026**

For further enquiry, please contact HKIMLSQAP Ltd.
Phone: (852) 2499 0015 Fax: (852) 2124 2798 E-mail: info@hkimlsqap.org

CONFIDENTIALITY

HKIMLSQAP is committed to keep all details of participants confidential. Please refer to http://www.hkimlsqap.org

HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES
QUALITY ASSURANCE PROGRAMME
ANATOMICAL PATHOLOGY

AUGUST SURVEY (2026)

HISTOLOGICAL STAINING MODULE

You are provided with two paraffin sections, which had been fixed in 10% buffered formalin. Perform the following staining methods and return the **STAINED SECTIONS** for assessment.

HC2611 Stain with your routine H&E method

HC2612 Stain with Masson's Trichrome method to demonstrate connective tissue

POSITIVE CONTROLS ARE REQUIRED BUT NOT FOR ASSESSMENT.
Please label the slides with the corresponding **Test code** and your confidential **Laboratory Code** for identification purposes. **No mark will be given for the unlabelled slide.** Put not more than two slides in one mailer in each return.

HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES
QUALITY ASSURANCE PROGRAMME

HISTOLOGICAL STAINING MODULE
RETURN FORM

Laboratory Code: | | | | | Date of Return: on or before **20 August 2026**

QUESTIONNAIRE

SECTION I: HISTOPATHOLOGY

1. What is your routine fixative for connective tissue stain?

   10% buffered formalin _____
   Bouin's fluid _____
   Zenker's fluid _____
   Others (please state) _____

2. What is the section thickness for your routine connective tissue stain?

   3 µm _____
   4 µm _____
   5 µm _____
   Others (please state) _____

3. What is your routine stain to demonstrate connective tissue?

   Masson trichrome _____
   Mallory trichrome _____
   Van Gieson _____
   Others (please state) _____

4. What other staining method(s) you will routinely use to demonstrate connective tissue?

   None _____
   Van Gieson _____
   Mallory trichrome _____
   Gomori's one step trichrome _____
   Others (please state) _____

5. Do you mordant the section before Masson's Trichrome staining?
   Yes _____
   No _____
   If "yes", please state the mordant and the duration you use.
   _____

6. What do you use for the differentiation?
   PMA _____
   PTA _____
   Others (please state) _____
   _____

7. What iron haematoxylin do you use for nuclear staining?
   Weigert' iron haematoxylin _____
   Celestine Blue – Mayer's haematoxylin _____
   Others (please state)
   _____

8. What counterstain do you use for Masson's trichrome stain?
   Light green _____
   Methyl Blue _____
   Others (please state)
   _____

9. Your routine histochemical procedure:

Masson's Trichrome Method

Reagent	***%/Compostion***	***Time***	***Temp(^{o}C)***
Haematoxylin		min	
Ponceau Fuchsin	Ponceau Fuchsin ____ gm ____% HAc _____ml	min	
Differentiation	____% PMA _or ____% PTA or Others: _____	min.	
Counterstain	____% Light Green or ____% Methyl Blue or Others: _____	min	

END

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ANATOMICAL PATHOLOGY

AUGUST SURVEY (2026)

IMMUNOHISTOCHEMICAL STAINING MODULE

Evaluation of p53 Antibody

HC2613 You are provided with a paraffin section, stain the slide with your **in-house Anti-Human p53** antibody and your routine immunohistochemistry detection system.

Evaluation of TTF-1 Demonstration

HC2614 You are provided with a paraffin section, stain the slide with your **in-house Anti-Human TTF-1** antibody and your routine immunohistochemistry detection system.

Evaluation of CD117 Demonstration

HC2615 You are provided with a paraffin section, stain the slide with your **in-house Anti-Human CD117** antibody and your routine immunohistochemistry detection system.

POSITIVE CONTROLS ARE REQUIRED BUT NOT FOR ASSESSMENT.

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QUALITY ASSURANCE PROGRAMME

IMMUNOHISTOCHEMICAL STAINING MODULE
RETURN FORM

Laboratory Code: | | | | | Date of Return: on or before **20 August 2026**

QUESTIONNAIRE

Your immunohistochemistry conditions:

STEP	***HC2613 (p53 in-house)***	***HC2614 (TTF-1 in-house)***	***HC2615 (CD117 in-house)***
Supplier			
Dilution	1:	1:	1:
Peroxidase Blocking	min.	min.	min.
Antigen retrieval: □ Trypsinization □ Microwave □ Pressure cooking □ PT module □ Others	YES / NO min.	YES / NO min.	YES / NO min.
Detection System			
Duration of Colour Development	DAB min.	DAB min.	DAB min.
End product Colour Enhancement (if any)	min.	min.	min.

END